NITROGEN SATURATION IN STREAM ECOSYSTEMS

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Abstract. The concept of nitrogen (N) saturation has organized the assessment of N loading in terrestrial ecosystems. Here we extend the concept to lotic ecosystems by coupling Michaelis-Menten kinetics and nutrient spiraling. We propose a series of saturation response types, which may be used to characterize the proximity of streams to N saturation. We conducted a series of short-term N releases using a tracer $({}^{15}NO_3-N)$ to measure uptake. Experiments were conducted in streams spanning a gradient of background N concentration. Uptake increased in four of six streams as NO₃-N was incrementally elevated, indicating that these streams were not saturated. Uptake generally corresponded to Michaelis-Menten kinetics but deviated from the model in two streams where some other growth-critical factor may have been limiting. Proximity to saturation was correlated to background N concentration but was better predicted by the ratio of dissolved inorganic N (DIN) to soluble reactive phosphorus (SRP), suggesting phosphorus limitation in several high-N streams. Uptake velocity, a reflection of uptake efficiency, declined nonlinearly with increasing N amendment in all streams. At the same time, uptake velocity was highest in the low-N streams. Our conceptual model of N transport, uptake, and uptake efficiency suggests that, while streams may be active sites of N uptake on the landscape, N saturation contributes to nonlinear changes in stream N dynamics that correspond to decreased uptake efficiency.

Key words: Michaelis-Menten; nitrate; nitrogen spiraling; nitrogen uptake; saturation; stable isotope; streams.

INTRODUCTION

Nutrient input to terrestrial landscapes is leading to increased nitrogen (N) loading to aquatic systems. Headwater streams are important links in the transport of N from enriched terrestrial systems, but in-stream biological processes may also reduce the downstream flux of N (Alexander et al. 2000, Peterson et al. 2001, Bernhardt et al. 2003, Hall 2003). At the same time, the abundance and diversity of stream biota may respond to altered N supply over short- or long-term time scales. Aber et al. (1989, 1998) proposed the N-saturation hypothesis whereby enhanced inputs of N outpace forest uptake. They describe a series of N-saturation stages characterized by distinct, nonlinear changes in N processing leading to increased N loss at the highest stages of N saturation. By analogy to forests, saturated low-order streams should pass higher N loads on to downstream reaches. Here we present an analysis of the relationship between stream water N concentration and uptake in stream ecosystems spanning a range of chronic N availability. We used a stable isotope tracer (^{15}N) to determine how N uptake by streams spanning a gradient of dissolved inorganic N (DIN) concentration responds to experimental increases in N concentration. We

combined the nutrient spiraling concept (Newbold et al. 1981) to a Michaelis-Menten (M-M) framework to identify and characterize saturation response types analogous to the stages of N saturation described by Aber et al. (1989, 1998) for forests.

N availability, uptake kinetics, and saturation response types

The M-M model (Eq. 1) was originally developed to model enzyme kinetics but has proven to be robust in describing nutrient uptake as a function of concentration at a broad range of scales:

$$U = \frac{U_{\max}C}{C + K_{\max}} \tag{1}$$

where U is uptake, C is nutrient concentration, $K_{\rm m}$ is the half-saturation constant, and $U_{\rm max}$ is maximum uptake. Maximum uptake and the half-saturation constant ($K_{\rm m}$, the concentration at which uptake is one-half of $U_{\rm max}$) vary widely among organisms and in response to environmental conditions. These metrics are indices of organismal or system affinity for a nutrient.

At low concentrations (i.e., $C \ll K_m$), uptake approximates a linear relationship with increasing nutrient concentration (Kim et al. 1990). As nutrient concentration increases to near-saturating concentrations, U approaches U_{max} asymptotically. We contend that these regions of the M-M curve can be abstracted to represent three distinct stages of N saturation in stream ecosystems. The response of uptake in a stream to short-

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Stream	Location		NH ₄ -N (µg/L)	SRP (µg/L)	N:P	Width (cm)	Depth (cm)	Discharge (L/s)
Sammy Creek I (spring)	Giles County, Virginia	5	5	bdl	3	136	5	7
Sammy Creek II (summer)	Giles County, Virginia	3	bdl	bdl	2	71	4	6
Hugh White Creek	Macon County, North Carolina	3	3	bdl	2	225	2	7
Alta Creek	Montgomery County, Virginia	182	bdl	7	25	249	8	15
Stonecrop Creek	Giles County, Virginia	298	5	20	16	101	3	2
Greenbrier Creek	Roanoke County, Virginia	983	bdl	5	195	190	8	31

TABLE 1. Characteristics of study streams used to assess N saturation.

Notes: Nitrate-nitrogen (NO₃-N), ammonium-nitrogen (NH₄-N), and soluble reactive phosphorus (SRP) are reach-average concentrations prior to all releases. N:P represents atomic ratios derived from the combined concentrations of NO₃-N and NH₄-N to SRP. The lowest detectable limit ($3 \mu g/L$) was used to calculate N:P when measured concentrations were below detectable limits (bdl). Width, depth, and discharge reflect reach mean measurements.

term additions of N should follow one of these saturation response types (SRTs) that correspond to a stream's proximity to uptake saturation. Thus, SRTs are delineated by the relationship between uptake and concentration, including: (1) Type I, exhibiting a constant, linear increase (slope > 0) in uptake, indicating that a stream is well below saturation, (2) Type II, uptake increasing to an asymptotic plateau (curvilinear, slope > 0 but declining), indicating that a stream is approaching saturation, and (3) Type III, no increase in uptake in response to nutrient amendment, characterizing a nutrient-saturated stream.

Linking uptake kinetics, N saturation, and nutrient spiraling

Solutes in streams are simultaneously affected by processes that assimilate or transform nutrients and by forces that cause downstream transport of dissolved and suspended material. The nutrient spiraling concept (Webster and Patten 1979) incorporates both downstream transport and cycling of nutrients within streams and provides a conceptual and technical framework for stream biogeochemistry (Newbold et al. 1981). Three commonly measured indices are used to describe spiraling in streams. (1) Uptake length (S_w) is the mean distance traveled as a dissolved inorganic solute before immobilization and removal from the water column. (2) Uptake velocity (v_f) represents the theoretical speed at which a nutrient moves toward a sink (i.e., stream bottom) and standardizes for stream depth and velocity (Stream Solute Workshop 1990):

$$v_{\rm f} = \frac{d}{S_{\rm w}} u \tag{2}$$

where u is stream-water velocity and d is stream depth. (3) Areal specific uptake (U) quantifies the mass of nutrient immobilized per area of streambed per unit time (Stream Solute Workshop 1990):

$$U = v_{\rm f}C\tag{3}$$

where C is stream water nutrient concentration. Examination of Eqs. 2 and 3 shows that, in addition to standardizing for stream morphometry and hydraulics, $v_{\rm f}$ reflects nutrient uptake relative to concentration

(i.e., U/C) and thus serves as a descriptor of nutrient uptake efficiency (Davis and Minshall 1999).

Because the spiraling metrics (S_w , v_f , and U) are mathematically related, applying the M-M model to Udictates how S_w and v_f should respond to increasing nutrient concentration. The uptake velocity (v_f) is described by a nonlinear decrease with increasing nutrient concentration:

$$v_{\rm f} = \frac{U_{\rm max}}{K_{\rm m} + C}.\tag{4}$$

At the same time, S_w increases linearly with increasing nutrient concentration:

$$S_{\rm w} = ud\left(\frac{C+K_{\rm m}}{U_{\rm max}}\right).$$
 (5)

Both $K_{\rm m}$ and $U_{\rm max}$ can be derived from empirical relationships between nutrient concentration and any of the spiraling metrics. We used stable N-isotope (i.e., 15 N) solute release experiments to assess ecosystem uptake response to experimental N additions in streams spanning a gradient of N concentration.

Methods

Study sites

We examined N uptake across five first- or secondorder headwater streams in the southern Appalachian Mountains of southwestern Virginia and western North Carolina, USA (Table 1) selected to span a gradient of background dissolved inorganic nitrogen (DIN) concentration. The concentration of DIN in our study streams is characteristic of the range of DIN concentration reported in other streams throughout the region (McTammany 2004, Hagen et al. 2006; Webster and Valett, unpublished data). Streams were bordered by forested riparian zones, but land use within associated catchments was variable. Two streams (Sammy Creek and Hugh White Creek) were located within forested catchments that have been relatively undisturbed by human activities since the 1920s. Vegetation included mixed deciduous forests with thick streamside canopies of rhododendron (Rhododendron maxium L.). Catchments of the other streams (Alta Creek [see Plate 1], Greenbrier Creek, and Stonecrop Creek) included a

TABLE 2. Enrichment levels and spiraling metrics from individual releases.

Stream and date	Discharge (L/s)	NO ₃ -N (µg/L)	S _w (m)	(mm/min)	$\bigcup_{(\mu g \ N \cdot m^{-2} \cdot min^{-1})}^{U}$		
Sammy Creek I							
29 April 2003 1 May 2003	9.0 6.8	5 47	$128 \pm 21 \\ 181 \pm 22$	1.57 0.83	7.8 39.0		
Sammy Creek II							
15 July 2003 16 July 2003	6.1 6.1	3 140	$42 \pm 3 \\ 88 \pm 9$	2.09 0.74	5.1 103.7		
Hugh White Creek							
8 October 2003 9 October 2003 10 October 2003 11 October 2003	6.8 7.0 7.7 8.2	3 39 260 537	$\begin{array}{r} 23 \pm 2 \\ 175 \pm 2 \\ 508 \pm 14 \\ 816 \pm 15 \end{array}$	2.80 0.37 0.14 0.09	8.8 14.6 36.8 48.1		
Alta Creek							
25 August 2003 26 August 2003 27 August 2003 28 August 2003	15.5 15.3 15.6 13.5	181 223 360 577	$\begin{array}{c} 587 \pm 81 \\ 1157 \pm 135 \\ 3128 \pm 543 \\ 5464 \pm 715 \end{array}$	0.46 0.22 0.08 0.05	84.0 49.8 30.2 26.8		
Stonecrop Creek							
 September 2003 September 2003 September 2003 September 2003 	2.2 2.0 1.9 2.1	287 361 575 850	632 ± 24 684 ± 84 1002 ± 38 1263 ± 131	$0.09 \\ 0.09 \\ 0.06 \\ 0.05$	26.5 30.7 35.7 41.8		
Greenbrier Creek 14 August 2003	32.0	981	3652 ± 1037	0.23	222.4		
14 August 2003 15 August 2003 15 August 2003	32.0 29.0 29.0	1081 1240 1568	5519 ± 1264 7194 ± 1136	0.14 0.11	173.1 167.9		

Notes: Values for uptake length (S_w) are means \pm se. NO₃-N represents the geometric mean of the NO₃-N concentration (μ g/L) in the study reach during plateau sampling. NS indicates a nonsignificant ($P \ge 0.05$) regression.

mixture of forest, small farms, and residential areas. The substratum of Sammy Creek and Alta Creek was dominated by small cobbles and boulders, whereas a mix of sand, gravel, and silt dominated the substratum at each of the other three streams (Earl 2004). Study streams were characterized by soft water, with the exception of Stonecrop Creek, which drained a predominately limestone catchment. We selected a single study reach (50–388 m) in each stream, with length depending on accessibility, stream water travel time, and distance appropriate to accurately measure N uptake.

Solute addition experiments

A series (2–4) of short-term (3–5 h) nutrient releases were used to manipulate stream water nitrate-N (NO₃-N) concentration. At the same time, $^{15}NO_3$ -N was added as a tracer to measure uptake response to increasing NO₃-N availability. At each stream, a single release was conducted each day over several consecutive days except at Greenbrier Creek, where two releases were conducted consecutively over two days (Table 2). Releases were conducted during periods without storms to address generally similar climatic and hydraulic conditions. Prior to release, background samples were collected from four to seven points along the study reach and analyzed for NO₃-N, chloride (Cl[¬]), and ¹⁵NO₃-N. A subset of the background samples was analyzed for ambient concentrations of ammonium-N (NH₄-N) and soluble reactive phosphorus (SRP).

For the first release, a solution of K¹⁵NO₃ (98 atom percentage; Cambridge Isotope Laboratories, Andover, Massachusetts, USA) and NaCl (Cl⁻ to act as a conservative tracer, Bencala et al. 1990) was released at a constant rate designed to increase stream water δ^{15} NO₃-N by 500‰. Tracer ¹⁵NO₃-N additions elevated stream water NO₃-N concentration 11% above ambient at Hugh White Creek, and 9% and 3% at Sammy Creek II (summer) and I (spring), respectively, and <0.6% at all other sites.

Three replicate water samples were collected from four to seven points (depending upon study reach length) downstream of the release site under well-mixed conditions (i.e., as indicated by steady-state conductivity in channel water). Samples for the analysis of ¹⁵NO₃-N were collected in acid-washed 4-L bottles, chilled until filtered (glass-fiber filters, Whatman GF/F, pore size = 1.0 µm; Florham Park, New Jersey, USA), and refrigerated (~4°C) for less than one week until processed. Additional samples (n = 3 per transect) were collected for analysis of NO₃-N and CL⁻, filtered (glassfiber filters, Gelman A/E, pore size = 1.0 µm; Ann Arbor, Michigan, USA), and frozen that day. Identical methods were employed for subsequent ¹⁵N releases at each stream; however, varying quantities (Table 2) of NaNO₃ were added to the release solution to increase stream water NO₃-N concentration. Target $\delta^{15}NO_3$ -N (i.e., 500‰) was maintained by adjusting the amount of K¹⁵NO₃ in the release solution as necessary.

Stream water conductivity was continuously monitored at the downstream end of the study reach, and velocity was determined from the corresponding conservative tracer curve (Bencala and Walters 1983). Discharge (Q) at each transect was determined by dilution gauging using the conservative tracer (Cl[¬]) and assumed to be constant throughout sampling. Widths and depths of the wetted channel were measured at 5–10 m intervals along the study reach prior to or following the series of releases.

Laboratory methods

Anions (NO₃-N and Cl⁻) were analyzed on a Dionex DX500 Ion Chromatograph (Dionex, Sunnyvale, California, USA). Due to low NO₃-N concentrations (<10 μ g/L), samples collected at Sammy Creek and Hugh White Creek were analyzed colorimetrically following reduction by Cd (Wood et al. 1967, APHA 1998) on a Technicon Autoanalyzer (Technicon, Emeryville, California, USA). Samples were analyzed for NH₄-N using a modified phenol–hypochlorite method (Solorzano 1969, USEPA 1997*a*) and SRP using the molybdate–antimony method (Murphy and Riley 1962, USEPA 1997*b*). DIN was calculated as the sum of NO₃-N and NH₄-N. Atomic N:P ratios were determined using DIN and SRP concentrations.

¹⁵NO₃-N was measured by headspace diffusion according to Sigman et al. (1997). Ammonium was removed by boiling samples to a final volume of ~ 100 mL under basic conditions by adding MgO. Nitrate-N was converted to ammonia (NH₃) by adding Devarda's Alloy. A precombusted, acidified (25 µL, 2.5 M KHSO₄) glass-fiber filter (Whatman GF/D) encased by Teflon tape was added to the sample immediately after adding the Devarda's alloy. Sealed samples were incubated at 60°C for 48 h and then shaken gently for 10 d during which time NH₃ was captured by the acidified filter. The filters were then removed from the Teflon sandwich, dried, tinned, and shipped to the University of California, Davis Stable Isotope Facility (Davis, California) for analysis of ¹⁵N on a Europa Integra mass spectrometer (Sercon, Cheshire, UK).

In the two streams with background NO₃-N concentrations <80 µg/L (Sammy Creek and Hugh White Creek; Table 1), background and plateau samples were spiked with a known amount of NO₃-N to ensure a minimum mass (\geq 80 µg) of N required to measure ¹⁵NO₃-N in a reasonable sample volume (i.e., \leq 1 L). Thus, the target δ ¹⁵NO₃-N of the field release was elevated such that the sample δ ¹⁵NO₃-N would reflect an enrichment of 500‰ after spike addition.

Data analysis

Uptake length (S_w) was calculated by fitting an exponential decay model (SigmaPlot, SPSS, Chicago, Illinois, USA) to the background- and dilution-corrected flux of ¹⁵NO₃-N (determined following Mulholland et al. 2000) vs. distance downstream. The decay coefficient represents the longitudinal loss rate (k_L), and its negative inverse is the uptake length (Newbold et al. 1981). The standard error (SE) of S_w was determined as the standard error of the uptake coefficient using a modified exponential decay model:

¹⁵NO₃-N flux =
$$ae^{(x/S_w)}$$
. (6)

where a is equal to the ¹⁵NO₃-N flux at the upper-most transect, and x is distance downstream. Uptake velocity $(v_{\rm f})$ and uptake (U) were calculated according to Stream Solute Workshop (1990) following Eqs. 2 and 3, respectively. In Eq. 3, C was the geometric mean of plateau NO₃-N from all sampling points. Linear and nonlinear regression were used to relate spiraling metrics $(S_{\rm w}, v_{\rm f}, U)$ to NO₃-N amendment concentration. Linear regression, performed with SigmaStat (SPSS, Chicago, Illinois, USA), was used to assess the relationship between S_w and NO₃-N amendment concentration. The nonlinear regression function in SigmaPlot was used to relate amendment NO₃-N concentration to $v_{\rm f}$ and U according to Eqs. 4 and 1, respectively. In addition, whole-stream estimates of K_m and U_{max} were calculated from each of these relationships. The slopes of the relationships between Sw and NO₃-N amendment for each stream were compared by ANCOVA using the PROC REG procedure in SAS. Significance level (α) was set to 0.05 for all statistical tests.

Saturation response type (SRT) for each stream was determined by the best-fit regression between U and NO₃-N amendment concentration. The relationship at each stream was assessed with a linear and nonlinear (Eq. 1) model. A significant linear model indicated an SRT I designation whereas a significant nonlinear model with positive coefficients indicated an SRT II designation. In the event that both models were statistically significant, model r^2 was used to identify the appropriate model. Streams were designated SRT III if there was no statistically significant increase in uptake. Model significance and r^2 were used to identify the best-fit model in lieu of traditional goodness-of-fit tests, due to the small sample size.

RESULTS

All five streams were relatively small, with mean depth ranging between 2 cm and 8 cm, and mean width ranging between 71 cm and 249 cm (Table 1). The study streams varied in discharge from 2 L/s to 31 L/s at the time of the release, and mean concentrations of NH₄-N were consistently low ($\leq 5 \ \mu g/L$), while background NO₃-N varied from 3 $\mu g/L$ to 982 $\mu g/L$ (Table 1). The concentration of SRP was $\leq 7 \ \mu g/L$, with the exception



FIG. 1. Response of uptake length (S_w , mean \pm sE), the distance traveled as a dissolved inorganic solute before immobilization and removal from the water column, to N amendments for all releases at Sammy Creek I, Sammy Creek II, Hugh White Creek, Alta Creek, Stonecrop Creek, and Greenbrier Creek. The first point for each stream corresponds to S_w measured at the ambient stream water NO₃-N concentration; subsequent points correspond to S_w measured in association with NO₃-N amendments. Nitrogen amendment concentration is presented as the geometric mean of plateau NO₃-N concentration (μ g/L) in the study reach. Linear regression lines are presented for all streams except Sammy Creek I and II, for which only two releases were conducted during each experiment.

of Stonecrop Creek (20 μ g/L). Atomic N:P varied from 2 to 195 among sites.

Response to N amendments

For all streams, uptake length (S_w) increased with increasing NO₃-N amendment (Fig. 1). Significant, positive, linear relationships between S_w and NO₃-N concentration occurred at Alta Creek, Stonecrop Creek, and Hugh White Creek ($r^2 \ge 0.975$, $P \le 0.013$). While S_w increased with NO₃-N amendment concentration during experiments at Sammy Creek (I and II), only two additions were completed at these sites, thus limiting curve assessment. Uptake length also increased with increasing NO₃-N amendment concentration at Greenbrier Creek; however, the relationship was not significant ($r^2 = 0.990$, P = 0.063), reflecting low statistical power (i.e., n = 3 releases). The relationship between ¹⁵NO₃-N flux and distance downstream during the first amended release at Greenbrier Creek (14 August 2003, 1081 µg NO₃-N/L; Table 2) was not significant (P =0.141) resulting from unusually high variability of $\delta^{15}NO_3$ -N values. This was the only occurrence of a nonsignificant estimate of S_w , and the value was not plotted or included in analyses. In general, there was considerable uncertainty surrounding estimates of S_w at Greenbrier Creek (Table 2, Fig. 1).

Uptake velocity (v_f) declined in a curvilinear manner with increasing NO₃-N amendment concentration at most sites (Fig. 2). Significant ($P \le 0.029$) relationships between v_f and NO₃-N amendment concentration were evident at Alta Creek, Stonecrop Creek, and Hugh White Creek. Similarly, v_f declined in a curvilinear manner at Greenbrier Creek, but the regression was not significant (P = 0.065, Fig. 2). In addition, $v_{\rm f}$ decreased with NO₃-N amendment concentration during both spring and summer collections at Sammy Creek.

Relationships between uptake (U) and NO₃-N amendment concentration differed among sites (Fig. 3). At Sammy Creek, U increased with increasing NO₃-N amendment concentration during both experiments. Despite an apparent curvilinear fit ($r^2 = 0.940$, Eq. 1), the relationship between U and NO₃-N amendment concentration at Hugh White Creek was best described by a linear model ($r^2 = 0.947$). Uptake and NO₃-N amendment concentration at Stonecrop Creek was better fit by an asymptotic model ($r^2 = 0.989$) than a linear model ($r^2 = 0.976$). Uptake generally declined with NO₃-N amendment concentration at Greenbrier Creek and Alta Creek, but linear regression indicated that the slope of the relationship between U and NO₃-N amendment concentration was not significant for either stream (Fig. 3).

For a given stream, estimates of $K_{\rm m}$ and $U_{\rm max}$ calculated from each of the spiraling metrics were generally similar (coefficient of variation [mean/sD] \leq 52% for all but Hugh White Creek; Table 3). Differences among values of $K_{\rm m}$ or $U_{\rm max}$ calculated from each of the three spiraling metrics within a given stream likely reflect variability in variables associated with conversion (i.e., u, d, and C). Over four of the six experiments, mean $K_{\rm m}$ varied from 59 to 303 µg/L and $U_{\rm max}$ from 19 to 158 µg N·m⁻²·min⁻¹ (Table 3). All calculated values for $K_{\rm m}$ were negative, and calculated $U_{\rm max}$ values were far



FIG. 2. Uptake velocity (v_f) responses to N amendments. The first point in each panel corresponds to v_f measured at the ambient stream water NO₃-N concentration; subsequent points correspond to v_f measured in association with NO₃-N amendments. Nitrogen amendment concentration is presented as the geometric mean of plateau NO₃-N concentration ($\mu g/L$) in the study reach. Results of nonlinear regression following Eq. 4 are presented for those streams for which more than two releases were conducted.

below observed uptake (U) at both Greenbrier Creek and Alta Creek (Table 3).

Negative estimates of M-M parameters (i.e., K_m and U_{max}) at Greenbrier Creek and Alta Creek reflect basic differences in spiraling responses to N amendment compared to the other study streams. The negative intercepts associated with the regression relating S_w and NO₃-N amendment at these two streams corresponded to the negative K_m values (Eq. 5, Table 3). Slopes of S_w vs. NO₃-N amendment concentration at both Greenbrier Creek and Alta Creek were significantly greater (ANCOVA, $P \leq 0.002$, $\alpha = 0.008$, after Bonferroni correction) than slopes at both Stonecrop Creek and Hugh White Creek. Due to the limited number of data points, experiments at Sammy Creek were not included in the statistical comparison, but slopes from these

experiments were similar to those at Stonecrop Creek and Hugh White Creek (Fig. 1).

Patterns across streams

Spiraling metrics across study streams under ambient N conditions (i.e., ¹⁵NO₃-N tracer-derived) varied considerably depending on background stream-water N concentration. Ambient S_w increased from 23 m to 3652 m with increasing N concentration and a linear model explained most of the variability ($r^2 = 0.979$, P < 0.001). Ambient v_f varied from 0.09 mm/min to 2.80 mm/min and, as within streams, decreased nonlinearly (Eq. 2) with increasing background stream water NO₃-N concentration ($r^2 = 0.887$, P = 0.005). Ambient U varied from 5.1 µg·m⁻²·min⁻¹ to 222.4 µg·m⁻²·min⁻¹ and the relationship between U and the background stream



FIG. 3. Uptake (U) response to N amendments. Symbols and analysis details are as in Fig. 2. Regression models are either linear or curvilinear (following Eq. 1), depending on best fit as indicated by model r^2 .

water NO₃-N concentration was well described by both linear ($r^2 = 0.767$, P = 0.022) and nonlinear (Eq. 1; $r^2 = 0.902$, P = 0.004) models. Although regression statistics from the nonlinear model indicated a good fit to the

data, the model resulted in nonsensical coefficients ($K_{\rm m} > 1.9 \times 10^6 \,\mu g/L$, $U_{\rm max} > 4 \times 10^5 \,\mu g \cdot m^{-2} \cdot min^{-1}$), and a visual inspection indicated that the relationship was better fit by a linear model.

TABLE 3. Half-saturation constant (K_m) and maximum uptake (U_{max}) values calculated from regressions among NO₃-N amendments, S_w , v_f , and U, at five study streams from April through October 2004.

		$K_{\rm m}$ (µg/L)				$U_{\rm max} \ (\mu g \ { m N} \cdot { m m}^{-2} \cdot { m min}^{-1})$					
Stream	$S_{ m w}$	$v_{\rm f}$	U	Mean	CV	$S_{ m w}$	$v_{\rm f}$	U	Mean	CV	
Sammy Creek I†	96	42	42	60	52	138	74	74	95	39	
Sammy Creek II†	118	73	73	88	30	228	158	158	181	22	
Hugh White Creek	57	3	117	59	97	48	16	57	40	53	
Alta Creek	-126	-142	-141	-136	-7	21	18	19	19	8	
Stonecrop Creek	254	325	330	303	14	52	57	57	55	5	
Greenbrier Creek	-354	-503	-482	-446	-18	133	108	112	118	11	

 $\dagger K_{\rm m}$ and $U_{\rm max}$ values derived from two-point regressions.



FIG. 4. Cross-study comparison of NO₃-N spiraling across a N-concentration gradient. Regression results correspond to Eqs. 5, 4, and 1 for uptake length (S_w), uptake velocity (v_f), and uptake (U), respectively. Data sources are given in the figure; note that for Hall and Tank (2003), sites with infinite uptake lengths were excluded.

These patterns were evident, though less robust statistically, in a cross-stream, cross-study comparison of NO₃-N spiraling in streams throughout the coterminous United States that span a N gradient (Fig. 4). We observed statistically significant relationships between ambient NO₃-N concentration and uptake length ($r^2 = 0.386$, P < 0.001), uptake velocity ($r^2 = 0.075$, P = 0.043), and uptake ($r^2 = 0.436$, P < 0.001), following Eqs. 5, 4, and 1, respectively.

DISCUSSION

Biotic control of N uptake: whole stream measures

The relationships between S_w , v_f , U, and NO₃-N amendment corresponded to predicted patterns given M-M kinetics in four of six study streams. These results

suggest that biological processes are responsible for N uptake, and that uptake can be modeled according to M-M kinetics, but that alternate models of nutrient uptake may be more applicable in high-N streams. In contrast, Dodds et al. (2002) reported continual increase in N uptake, despite very high amendment concentrations in prairie streams. They proposed that the apparent lack of biotic saturation might have been due to hydrodynamic limitation across solid-water boundaries (i.e., mass transport limitation). Diffusion (i.e., mass transfer) of nutrients may be limited by benthic biofilm thickness, and increasing water velocity has been shown to stimulate growth of biofilms (Horner et al. 1990, Battin 2000). If uptake is limited solely by mass transport, uptake should be a linear function of



PLATE 1. Alta Creek, Montgomery County, Virginia (USA). Site of ¹⁵N releases in August 2003. Photo credit: S. Earl.

concentration, and both S_w and v_f would remain constant (Dodds et al. 2002). Biomass of benthic biofilms was low in these streams (Earl 2004), making it likely that biotic processes (i.e., uptake kinetics) were more important than diffusion-limited uptake in our streams.

Whole-stream estimates of $K_{\rm m}$ (3–330 µg/L, excluding Alta Creek and Greenbrier Creek) are generally within the range of those reported for other stream ecosystems and for freshwater algae. Whole-stream estimates of $K_{\rm m}$ ranged from 6 µg to 32 µg NH₄-N/L in prairie and mountain streams (Payn et al. 2005). Reported K_m values for freshwater algae include 59 µg NO₃-N/L for a periphyton assemblage (Kim et al. 1990), 1.4-130 µg N/ L for diatoms (USEPA 1985), and 102–213 μ g NO₃-N/ L for filamentous green alga (Cladophora glomerata, Lohman and Priscu 1992). General agreement between our estimates of K_m and those reported in the literature for micro- and mesocosm studies (where the influence of mass transport and adsorption is negligible) further supports biotic regulation of uptake in our study streams.

Saturation response types

The study sites spanned a continuum from potentially N-limited to N-saturated streams. An increase in uptake (U) with increasing N amendment at Sammy Creek,

Stonecrop Creek, and Hugh White Creek suggests that N uptake in these streams was not saturated. We observed evidence of Saturation Response Type (SRT) I at Hugh White Creek, and while the relationship between U and NO₃-N amendment at this site was best described by a linear model, visual inspection of the relationship suggested a slight nonlinear trend. This pattern may reflect a transition between SRTs I and II, or may result from the substantial range of NO₃-N amendment concentration used to address saturation in this stream. Very low background DIN concentration at Hugh White Creek resulted in amendments that elevated NO₃-N ~1- to 164-fold across the four releases while experimental enrichment varied from 1- to 58-fold at all other sites. Both experiments at Sammy Creek indicated that it was well below N saturation (SRT I). Although the relationship between U and NO3-N amendment could not be resolved due to the limited number of data points, uptake increased dramatically (a higher proportion of ambient U than in any other stream) despite a very high enrichment on both occasions. We observed evidence of SRT II at Stonecrop Creek, where U increased with increasing NO3-N amendment, according to an asymptotic relationship, and was approaching $U_{\rm max}$ at the highest amendment. Uptake did not increase significantly at Greenbrier Creek, corresponding to an SRT III and suggesting that uptake in this stream was saturated. Similarly, U did not increase significantly with increasing NO₃-N amendment at Alta Creek, suggesting that this system was also N saturated.

Based on our model, the relationship between U and NO3-N amendment at both Alta Creek and Greenbrier Creek denotes N saturation. However, visual inspection of the relationships suggests that U declined with increasing NO₃-N concentration at both sites, a trend not consistent with the M-M model. While the linear increase in S_w and nonlinear decline in v_f associated with NO₃-N augmentation at these sites correspond to predicted patterns given M-M kinetics, the mathematical relationships resulted in similar nonsensical estimates of $K_{\rm m}$ and $U_{\rm max}$. Further, the significantly greater slope of the relationship between S_w and NO₃-N amendment at both Alta Creek and Greenbrier Creek relative to all other sites suggests that these streams behaved differently than the other study streams with respect to N uptake.

These results suggest that alternate models of uptake of a given nutrient may be more applicable when streams are no longer limited by that nutrient (i.e., saturated). An increase in the availability of a nonlimiting nutrient should not alter uptake rates for that nutrient. Thus, the asymptotic relationship between U and nutrient concentration described by the M-M model may be truncated and, instead, should be modeled as zero-order (i.e., constant rate and slope of zero) as nutrient concentration increases beyond saturation (Dugdale et al. 1981). Traditionally, M-M model assessment addresses how reaction rates are limited by enzymatic kinetics. In real ecosystems, uptake and processing of one element may be influenced by the availability of another (Fairchild et al. 1985), and we contend that the transition in U from an asymptotic rise to a zero-order is illustrated by break points in other spiraling metrics as well. Specifically, the slope of the linear relationship between uptake length and concentration should be markedly steeper at concentrations in excess of saturation. At the same time, the transition to saturation should correspond to a greater rate of decline in nutrient uptake efficiency. Deviation of U from expected asymptotic increase, coupled with the significantly steeper slopes of the relationship between S_w and NO3-N amendment we observed at Alta Creek and Greenbrier Creek, correspond to expected patterns for systems exposed to N concentration in excess of saturation (i.e., model truncation). These observations suggest abrupt and nonlinear changes in N removal during stream transport with consequences for receiving waters. Such dramatic changes in N dynamics are central to the N-saturation hypothesis proposed for other ecosystems (Aber et al. 1998, Perakis et al. 2005).

The proximity of streams to N saturation, as indicated by their respective SRT was generally correlated to stream water nutrient concentration. Streams designated SRT I were low-N systems (Sammy Creek and Hugh White Creek). Our findings that low-N streams are not saturated concur with studies that reported N limitation at similar N concentrations. Benthic algae were N limited when concentrations were below 55 μ g NO₃-N/L in a desert stream (Grimm and Fisher 1986) and below 100 µg NO₃-N/L in an Ozark (USA) stream (Lohman et al. 1991). At the opposite end of the spectrum, streams designated SRT III (Alta Creek and Greenbrier Creek) had high N concentration. Our analysis indicated that Stonecrop Creek was approaching N saturation (i.e., characteristic of SRT II) but that the stream was not saturated, despite comparatively higher N concentration. Ultimately, nutrient saturation occurs when the availability of a nutrient increases to a point at which another factor critical to growth becomes limiting. In headwater stream ecosystems, nitrogen (Grimm and Fisher 1986), phosphorus (Hart and Robinson 1990), some combination of N and P (Winterbourn 1990), or other environmental factors (e.g., light; Lowe et al. 1986) often limit primary and secondary production. Lack of N saturation at Stonecrop Creek may reflect the high concentration of SRP in that stream, which resulted in a stream-water molar N:P ratio of 16. In a study of benthic algae, Shanz and Juon (1983) suggested that N was limiting at N:P ratios <10, P was limiting at N:P ratios >20, and the limiting nutrient was ambiguous at N:P ratios between 10 and 20. Nutrient ratios are useful for identifying limiting nutrients when concentrations are near growth-limiting concentrations (Borchardt 1996). Our analysis of N saturation in these study streams is consistent with the thresholds identified by Shanz and Juon (1983) and suggest that Stonecrop

Creek may not be N or P limited, whereas high N concentrations in Alta Creek and Greenbrier Creek may result in P limitation.

Stream response to chronic N loading

Nutrient uptake and biomass accrual are coupled only under steady-state conditions (Borchardt 1996), thus the uptake kinetics calculated from our short-term N amendments do not necessarily reflect kinetics that might be associated with long-term N loading within a given stream. However, N uptake and uptake efficiency are functions of biotic demand and nutrient concentration in all ecosystems. As such, spiraling metrics should respond in a characteristic fashion based on chronic N concentration. Predicted spiraling response across a gradient of N concentration translates into phases of N saturation analogous to the saturation response types (SRTs) described previously. We contend that, for streams experiencing low chronic levels of inorganic N, a restricted portion of the domain exists where the extant biota is able to use N subsidies in their entirety (Davis and Minshall 1999). Thus, phase I of N saturation in streams is described by short uptake lengths (S_w), high uptake efficiency (v_f), but relatively low uptake due to limited N availability. The N concentrations characteristic of phase I will depend on ambient standing stocks and diversity of stream biota responsible for primary uptake from the water column. Nevertheless, it appears that phase I is restricted to very low N concentrations. In Hugh White Creek, for example, despite a linear increase in uptake (U) as N increased from 3 to $>500 \mu g/L$, we observed an increase in uptake length (S_w) and nonlinear decline in uptake velocity (v_f) . While the stream was categorized as SRT I based on change in U, other metrics suggest that SRT II may be more appropriate for this range of N exposure. These responses suggest that streams may pass from phase I to phase II as ambient N increases only a very small amount. Further, this case illustrates how results from our assays of N saturation are sensitive to the range of N additions and their potential to alter patterns of ecosystem response. Accordingly, N saturation response curves must be interpreted in the context of the proposed extent of N enrichment. Uptake should be comparatively greater, uptake lengths longer, and uptake efficiencies lower (in accordance with the M-M model) in moderately enriched systems that correspond to phase II of N saturation. High-N systems (e.g., Greenbrier Creek and Alta Creek) indicative of the third phase of N saturation are characterized by relatively high uptake but markedly reduced uptake efficiency. At the same time, uptake lengths will be considerably longer in phase III as N atoms are transported increasingly longer distances before interaction with biotic sinks. Uptake did not change significantly, but Sw increased and $v_{\rm f}$ decreased dramatically at Greenbrier Creek, for Spiraling metrics under ambient N concentration in our study stream corresponded generally to predicted phases of N saturation. These patterns were also evident, though less robust, in a cross-study comparison of NO₃-N spiraling in disparate streams spanning a gradient of N concentration. While biotic processes are ultimately responsible for N cycling within streams (Hall and Tank 2003, Simon et al. 2005), these processes are influenced by a multitude of in-stream environmental variables, contributing to the considerable but expected variability within these relationships.

The high NO₃-N uptake efficiency in our low-N streams contributes to a growing body of evidence that documents the importance of streams as N-retention features on the landscape (e.g., Alexander et al 2000, Peterson et al. 2001, Bernhardt et al. 2003). However, our data show that the nature of N retention may change abruptly in response to N inputs. Regardless of uptake response, uptake efficiency declined nonlinearly with increasing experimental N amendment to streams. We also observed a pattern of decreasing uptake efficiency with higher background stream water N concentration (Fig. 4). These patterns are expected given M-M kinetics but also indicate that even unsaturated streams are less efficient retainers of N in the face of increased N loading. Annual rates of N loading into freshwater ecosystems has increased 6- to 50-fold since pre-industrial times, resulting in global fertilization in continental, estuarine, and near-coastal marine environments (Carpenter et al. 1998). Our analysis suggests that the capacity of streams to mitigate downstream fluxes of N is limited, potential retention or removal efficiency declines dramatically with even moderate inputs of N, and that streams may act to propagate N saturation down-gradient rapidly if N additions are not actively curtailed.

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